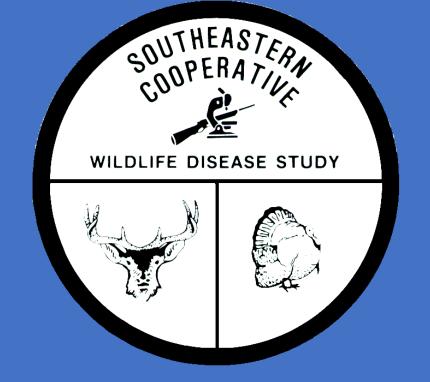
# Seeing what floats: Comparing fecal diagnostic techniques for the detection of zoonotic cestode eggs.

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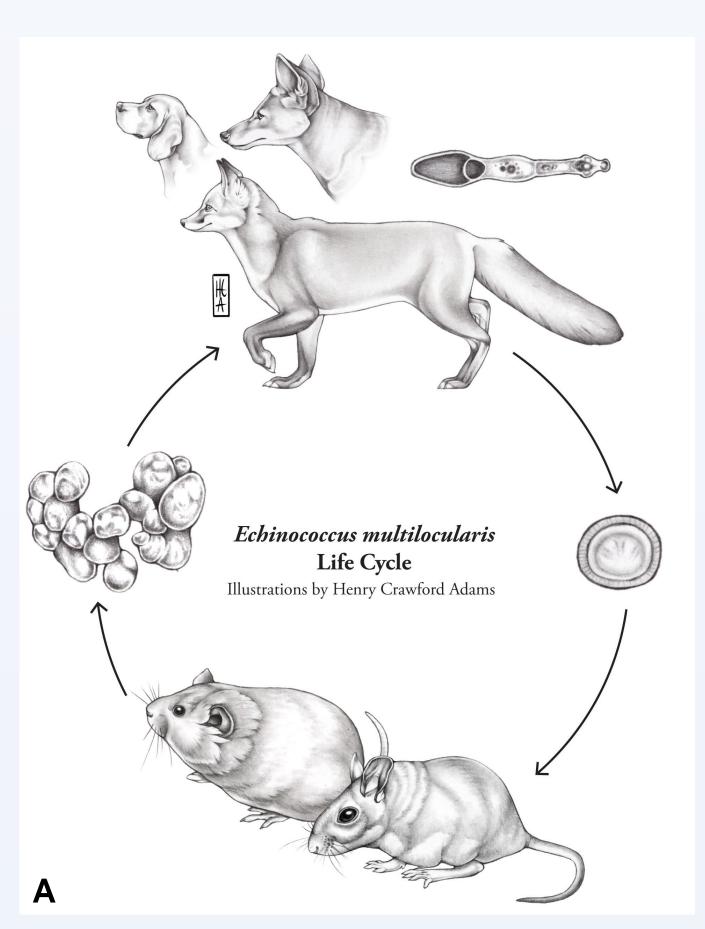
Sierra Felty<sup>1</sup>, Kayla Garrett<sup>2</sup>, Sarah Coker<sup>2</sup>, Ellen Haynes<sup>2</sup>, Michael Yabsley<sup>2</sup>, Christopher Cleveland<sup>2</sup>

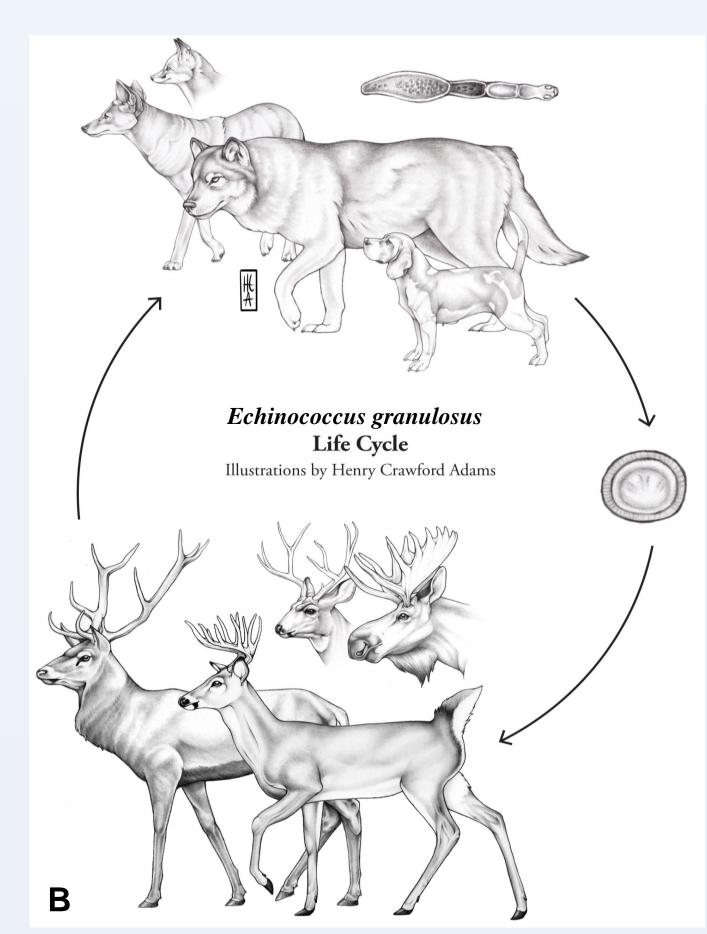


<sup>1</sup>Radford University <sup>2</sup> Southeastern Cooperative Wildlife Disease Study, University of Georgia

## INTRODUCTION

- Echinococcus species are parasitic cestodes (tapeworms) that infect humans, wildlife, livestock, and domesticated dogs and cats<sup>1</sup>.
- Definitive hosts are primarily canids, especially coyotes and foxes.<sup>1</sup>
- Reports of infection in non-endemic regions are increasing.<sup>2</sup>
- As urbanization increases worldwide, humans and domesticated animals are at greater risk of being infected through more frequent interactions with wild canid hosts.<sup>3</sup>
- Given the increased risk of transmission, there is a need to identify the most effective and sensitive fecal flotation methods to detect *Echinococcus* eggs.





**Figure 1.** Lifecycles of (A) *E. multilocularis* and (B) *E. granulosis*. Diagrams courtesy of Henry Adams.

# **RESEARCH OBJECTIVE & HYPOTHESIS**

- Different fecal flotation methods are employed by researchers and veterinary clinics, yet a *gold* standard test has not yet been widely adopted.
- The objective of this project was to compare the sensitivity & detection limit of three methods:
  - 1) Centrifugal
- 2) Passive
- 3) Mini-FLOTAC
- <u>Hypothesis</u>: Of the three methods, the centrifugal fecal float will be most sensitive for the detection of *Echinococcus* spp. eggs.

# **MATERIALS & METHODS**

- Isolated eggs from *Echinococcus* spp. proglottids
- Spiked negative fecal samples with known concentrations of *Echinococcus* spp. eggs: 25, 40, and 60 eggs per gram of feces.
- Performed 6 replicates of each method<sup>7,8,9</sup> using zinc sulfate flotation solution (1.18 SG):

## Centrifugal fecal float

- Homogenized 10mL flotation solution with 1g feces.
- Centrifuged for 10 minutes at 500 rpm then read.

## Passive fecal float

- Homogenized 1g of feces with 10mL flotation solution.
- Let sit in conical tube for 20 minutes then read.

## Mini-FLOTAC

- Homogenized 18mL flotation solution with 2g feces.
- Filled chambers, let sit 10 minutes, then read.

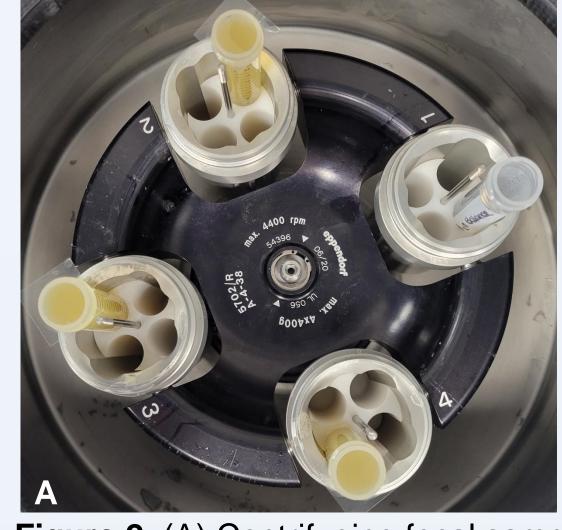




Figure 2. (A) Centrifuging fecal samples. (B) Echinococcus spp. egg.

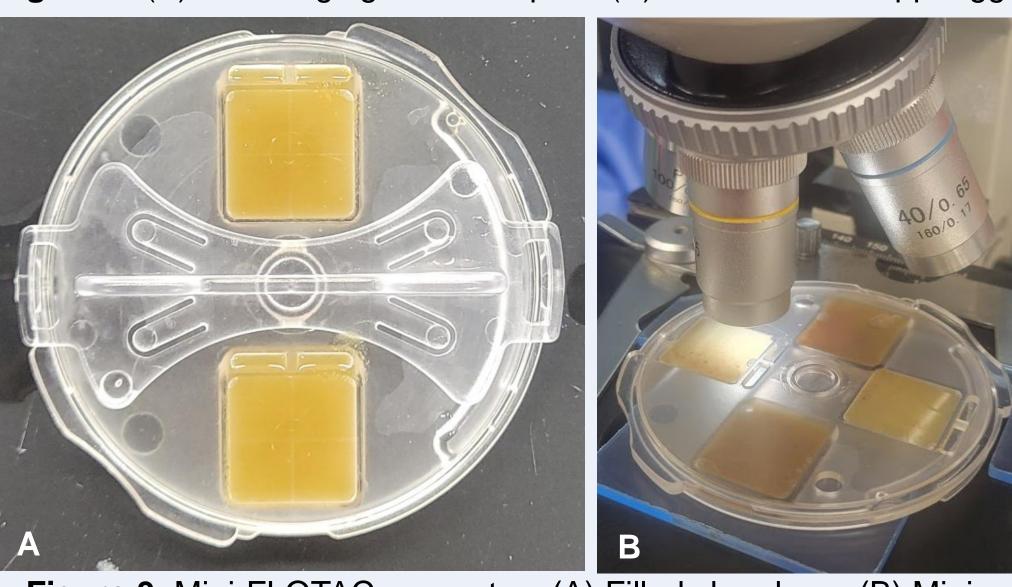
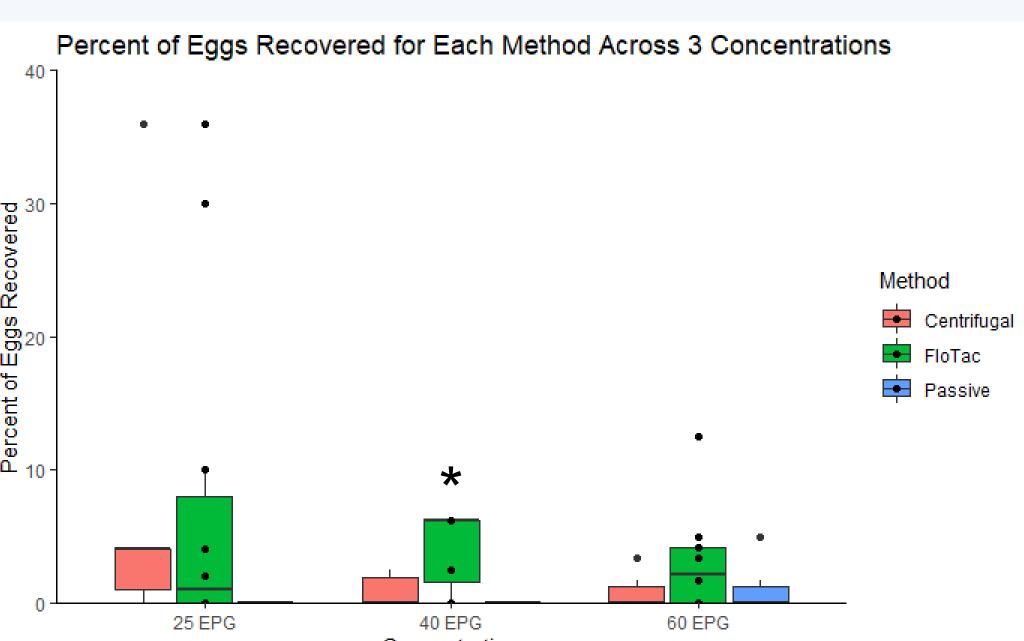


Figure 3. Mini-FLOTAC apparatus. (A) Filled chambers. (B) Mini-FLOTAC apparatus under microscope for reading.

# Percent of Positive Replicates for Each Method Across 3 Concentrations \* \*\* Method Centrifugal Flotac Passive

**Figure 4.** Percentage of replicates that were positive across three methods and concentrations. Asterisks denote statistically significant differences at a significance level of 0.05. EPG = eggs per gram.



**Figure 5.** Percent recovery of eggs across three different methods at differing concentrations. Asterisk denotes a statistically significant difference at a significance level of 0.05. EPG = eggs per gram.

## LIMITATIONS

- Eggs adhering to tools during sample preparation could result in lower concentrations.<sup>4</sup>
- Human error in collecting and counting eggs.
- Centrifugal fecal floats are prone to loss of coverslips during centrifugation.



Figure 6. Isolating Echinococcus sp. eggs.

## **CONCLUSIONS & SIGNIFICANCE**

- Centrifugal flotation and the Mini-FLOTAC had more positive results than passive flotation.
- The Mini-FLOTAC tended to detect more Echinococcus spp. eggs than the other methods.
- FLOTAC appears to be the best fecal flotation method to detect *Echinococcus* spp. eggs.
- Veterinarians in previously unaffected regions may not screen for *Echinococcus* spp.
- Public & veterinary health concern:
- Alveolar echinococcosis (AE) is particularly harmful to humans. Causative agent: E. multilocularis.<sup>5</sup>
- Cystic hydatid disease (CE) is less pathogenic. Causative agent: *E. granulosus.*<sup>5</sup>
- Echinococcus spp. currently infect 2-3 million people worldwide, costing \$200-800 USD million annually.<sup>6</sup>
- Infected livestock produce less meat, milk, and wool, resulting in an estimated \$1.5-\$2 billion USD loss annually.<sup>6</sup>
- Reliable detection methods are needed to address increasing *Echinococcus* spp. infections.

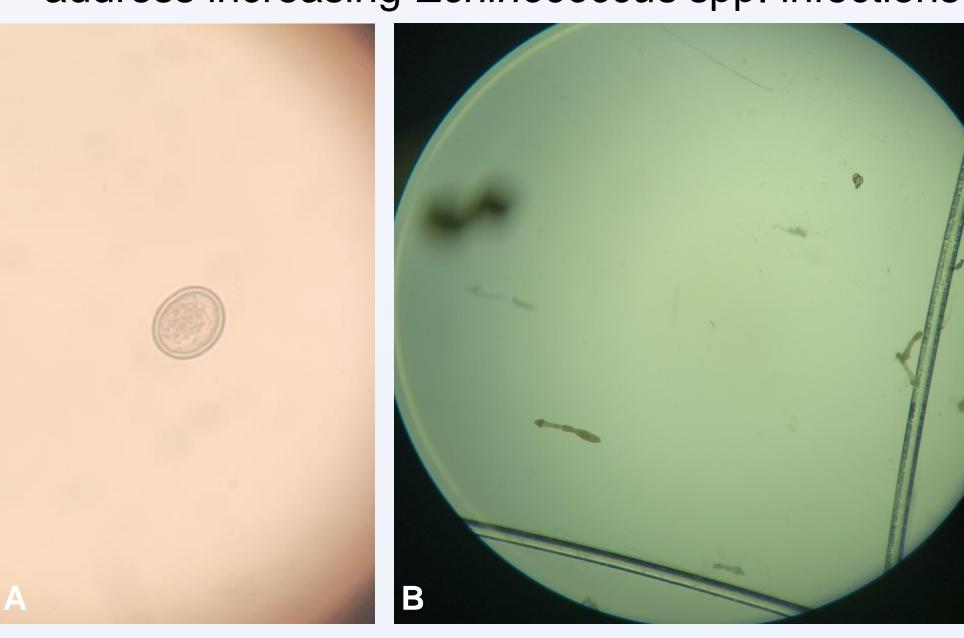


Figure 7. (A) Echinococcus spp. egg with distinct membrane. (B) Echinococcus spp. present in small intestinal sieve.

## **FUTURE DIRECTIONS**

- Continue data collection using larger sample sizes and different egg concentrations
- Co-author a manuscript detailing fecal flotation methods best suited for detection of *Echinococcus* spp. eggs.

## LITERATURE CITED

<sup>1</sup>Craig et al. *Echinococcus multilocularis*. 8 <sup>2</sup>Luong et al. (2020). Journal of Helminthology.

<sup>3</sup>Moro et al. (2009). International Journal of Infectious Diseases, 13(2).

<sup>4</sup>Kochanowski et al. (2014). Journal of Parasitology, 100(5).

Rochanowski et al. (2014). Journal of Parasi Roburer et al. (2014). Derecitalegy (144(2)

<sup>5</sup>Schurer et al. (2014). Parasitology, 141(2). <sup>6</sup>Cerda et al. (2018). Emerging Infectious Diseases, 24(2).

<sup>7</sup>Kida et al. (2022). Parasitology International.

<sup>8</sup>Kolapo et al. (2021). Veterinary Parasitology.

<sup>9</sup>Urquhart et al. (1996). Veterinary Parasitology (2<sup>nd</sup> edition).

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